

Instructions for Use

Chikungunya Virus IgM μ -capture ELISA

Enzyme immunoassay for the qualitative determination of IgM class antibodies against Chikungunya Virus in human serum or plasma.



1 x 96 well plate



ELS61243



Read the documentation

For Research Use Only

CONTENTS

1.	INTRODUCTION.....	3
2.	INTENDED USE	3
3.	PRINCIPLE OF THE ASSAY	3
4.	MATERIALS.....	3
4.1.	Reagents supplied.....	3
4.2.	Materials supplied	4
4.3.	Materials and Equipment needed.....	4
5.	STABILITY AND STORAGE	4
6.	REAGENT PREPARATION	4
6.1.	Coated Microplate	4
6.2.	Chikungunya Virus Antigen	4
6.3.	Washing Buffer (20x conc.)	4
6.4.	TMB Substrate Solution.....	4
7.	SAMPLE COLLECTION AND PREPARATION.....	5
7.1.	Sample Dilution	5
8.	ASSAY PROCEDURE	5
8.1.	Test Preparation.....	5
8.2.	Measurement	5
9.	RESULTS.....	6
9.1.	Run Validation Criteria.....	6
9.2.	Calculation of Results.....	6
9.3.	Interpretation of Results	6
10.	SPECIFIC PERFORMANCE CHARACTERISTICS	6
10.1.	Precision.....	6
10.2.	Diagnostic Specificity.....	7
10.3.	Diagnostic Sensitivity.....	7
10.4.	Interferences	7
10.5.	Cross Reactivity	7
11.	LIMITATIONS OF THE PROCEDURE.....	7
12.	PRECAUTIONS AND WARNINGS	7
12.1.	Disposal Considerations.....	7
13.	ORDERING INFORMATION.....	7
	BIBLIOGRAPHY.....	8
	ABBREVIATIONS.....	8
	SYMBOLS KEY	8

1. INTRODUCTION

Chikungunya virus is an arthropod borne virus of the genus Alphavirus (family Togaviridae). The Alphavirus genus contains at least 24 distinct species. These are lipid-enveloped virions with a diameter of 50 to 60 nm.

Alphavirus infections are initiated by the bite of an infected mosquito, which results in the deposition of virus in subcutaneous and possibly cutaneous tissues. After an incubation period of 1 to 12 days the Chikungunya fever develops.

Chikungunya fever (Chikungunya means “that which bends up”, in reference to the crippling manifestations of the disease) is an acute viral infection characterized by a rapid transition from a state of good health to illness that includes severe arthralgia and fever.

Temperature rises abruptly to as high as 40 °C and is often accompanied by shaking chills. After a few days, fever may abate and recrudescence, giving rise to a “saddleback” fever curve. Arthralgia is polyarticular, favoring the small joints and sites of previous injuries, and is most intense on arising. Patients typically avoid movement as much as possible. Joints may swell without significant fluid accumulations. These symptoms may last from 1 week to several months and are accompanied by myalgia. The rash characteristically appears on the first day of illness, but onset may be delayed. It usually arises as a flush over the face and neck, which evolves to a maculopapular or macular form that may be pruritic. The latter lesions appear on the trunk, limbs, face, palms and soles, in that order of frequency. Petechial skin lesions have also been noted. Headache, photophobia, retro-orbital pain, sore throat with objective signs of pharyngitis, nausea and vomiting also occur in this setting. Occasionally, however persistent arthralgia and polyarthritides (lasting months or even years) do occur, sometimes involving joint destruction. Even rarer, sequelae include encephalitis and meningoencephalitis with high lethality rates.

The virus has major importance in Africa and Asia. From 20% to more than 90% of the population of tropical and subtropical show serologic evidence of infection. Because Aedes mosquitoes are increasingly prevalent in North Africa and South America, where the population would be uniformly susceptible to infection, the possibility for epidemics is evident. Chikungunya virus infections are imported to central Europe mainly by travellers to tropical and subtropical countries.

Species	Disease	Symptoms (e.g.)	Transmission route
Chikungunya virus (Alphavirus)	Chikungunya fever	Fever; Exanthema; Joint pain Persistent arthralgia and polyarthritides, sometimes involving joint destruction. Even rarer encephalitis and meningoencephalitis.	Bite of insect vectors of the genus Aedes (Aedes albopictus, Aedes aegypti)

The presence of pathogen or infection may be identified by

- PCR
- Serology: by ELISA

2. INTENDED USE

The Chikungunya Virus IgM μ -capture ELISA is intended for the qualitative determination of IgM class antibodies against Chikungunya Virus in human serum or plasma (citrate, heparin).

3. PRINCIPLE OF THE ASSAY

The qualitative immunoenzymatic determination of specific IgM-class antibodies is based on the ELISA (Enzyme-linked Immunosorbent Assay) μ -capture technique.

Microplates are coated with anti-human IgM antibodies to bind the corresponding antibodies of the sample. After washing the wells to remove all unbound sample material, antigen is added. This antigen binds to the captured specific IgM antibodies. After a further washing step biotinylated antibody is pipetted into the wells. After washing horseradish peroxidase (HRP) labelled streptavidin is added that binds to the captured specific immune complex. After a further washing step the immune complexes are visualized by adding Tetramethylbenzidine (TMB) substrate which gives a blue reaction product. The intensity of this product is proportional to the amount of specific IgM antibodies in the sample. Sulphuric acid is added to stop the reaction. This produces a yellow endpoint colour. Absorbance at 450/620 nm is read using an ELISA microwell plate reader.

4. MATERIALS

4.1. Reagents supplied

- **Chikungunya Virus Coated Microplate (IgM):** 12 break-apart 8-well snap-off strips coated with anti-human IgM; in resealable aluminium foil.
- **Sample Diluent:** 1 bottle containing 100 ml of phosphate buffer (10 mM) for sample dilution; pH 7.2 \pm 0.2; coloured yellow; ready to use; white cap.
- **Stop Solution:** 1 bottle containing 15 ml sulphuric acid, 0.2 mol/l; ready to use; red cap.

- **Washing Buffer (20x conc.):** 2 bottles containing 25 ml of a 20-fold concentrated phosphate buffer (0.2 M), pH 7.2 ± 0.2, for washing the wells; white cap.
- **Chikungunya Virus antigen, lyophilized:** 6 bottles containing lyophilized Chikungunya virus antigen solution; red cap.
- **Chikungunya Virus antibody solution:** 1 bottle containing 6 ml of biotinylated Chikungunya virus antibody, ready to use; coloured blue; white cap.
- **Streptavidin conjugate:** 1 bottle containing 6 ml Streptavidin conjugated with peroxidase, ready to use; coloured red; black cap.
- **TMB Substrate Solution:** 1 bottle containing 15 ml 3,3',5,5'-tetramethylbenzidine (TMB), < 0.1 %; ready to use; yellow cap; < 5 % NMP. **Note: NMP may damage an unborn child. Wear protective gloves and eye protection when handling TMB Substrate Solution. Get medical advice in case of exposure.**
- **Chikungunya Virus IgM Positive Control:** 1 vial containing 1,5 ml control (human serum or plasma); coloured yellow; ready to use; red cap.
- **Chikungunya Virus IgM Cut-off Control:** 1 vial containing 2 ml control (human serum or plasma); coloured yellow; ready to use; green cap.
- **Chikungunya Virus IgM Negative Control:** 1 vial containing 1,5 ml control (human serum or plasma); coloured yellow; ready to use; blue cap.

For potential hazardous substances please check the safety data sheet.

4.2. Materials supplied

- 1 Cover foil
- 1 Instruction for use (IFU)
- 1 Plate layout

4.3. Materials and Equipment needed

- ELISA microwell plate reader, equipped for the measurement of absorbance at 450/620 nm
- Incubator 37 °C
- Manual or automatic equipment for rinsing wells
- Pipettes to deliver volumes between 10 and 1000 µl
- Vortex tube mixer
- Distilled water
- Disposable tubes

5. STABILITY AND STORAGE

Store the kit at 2...8 °C. The opened reagents are stable up to the expiry date stated on the label when stored at 2...8 °C.

6. REAGENT PREPARATION

It is very important to bring all reagents and samples to room temperature (20...25 °C) and mix them before starting the test run!

6.1. Coated Microplate

The break-apart snap-off strips are coated with anti-human IgM. Immediately after removal of the strips, the remaining strips should be resealed in the aluminium foil along with the desiccant supplied and stored at 2...8 °C.

6.2. Chikungunya Virus Antigen

The bottles contain lyophilized Chikungunya virus antigen solution. The content of each vial has to be resolved in 1 ml diluted Washing Buffer by turning it slowly (no vortex) and 15 min incubation at room temperature (20...25 °C). The reconstituted solution is stable for 1 day at 2...8 °C.

6.3. Washing Buffer (20x conc.)

Dilute Washing Buffer 1 + 19; e. g. 10 ml Washing Buffer + 190 ml distilled water. The diluted buffer is stable for 5 days at room temperature (20...25 °C). In case crystals appear in the concentrate, warm up the solution to 37°C e.g. in a water bath. Mix well before dilution.

6.4. TMB Substrate Solution

The reagent is ready to use and has to be stored at 2...8 °C, away from the light. The solution should be colourless or could have a slight blue tinge. If the substrate turns into blue, it may have become contaminated and should be thrown away.

7. SAMPLE COLLECTION AND PREPARATION

Use human serum or plasma (citrate, heparin) samples with this assay. If the assay is performed within 5 days after sample collection, the samples should be kept at 2...8 °C; otherwise they should be aliquoted and stored deep-frozen (-70...-20 °C). If samples are stored frozen, mix thawed samples well before testing. Avoid repeated freezing and thawing. Heat inactivation of samples is not recommended.

7.1. Sample Dilution

Before assaying, all samples should be diluted 1+100 with Sample Diluent. Dispense 10 µl sample and 1 ml Sample Diluent into tubes to obtain a 1+100 dilution and thoroughly mix with a Vortex.

8. ASSAY PROCEDURE

8.1. Test Preparation

Please read the instruction for use carefully **before** performing the assay. Result reliability depends on strict adherence to the instruction for use as described. The following test procedure is only validated for manual procedure. If performing the test on ELISA automatic systems we recommend increasing the washing steps from three to five and the volume of Washing Buffer from 300 µl to 350 µl to avoid washing effects. Pay attention to chapter 12. Prior to commencing the assay, the distribution and identification plan for all samples and standards/controls (duplicates recommended) should be carefully established on the plate layout supplied in the kit. Select the required number of microtiter strips or wells and insert them into the holder.

Perform all assay steps in the order given and without any delays.

A clean, disposable tip should be used for dispensing each standard/control and sample.

Adjust the incubator to 37 ± 1 °C.

1. Dispense 50 µl standards/controls and diluted samples into their respective wells. Leave well A1 for substrate blank.
2. Cover wells with the foil supplied in the kit.
3. **Incubate for 1 hour ± 5 min at 37±1°C.**
4. When incubation has been completed, remove the foil, aspirate the content of the wells and wash each well three times with 300 µl of Washing Buffer. Avoid overflows from the reaction wells. The interval between washing and aspiration should be > 5 sec. At the end carefully remove remaining fluid by tapping strips on tissue paper prior to the next step!
Note: Washing is important! Insufficient washing results in poor precision and false results.
5. Dispense 50 µl reconstituted Chikungunya virus Antigen into all wells except for the Substrate Blank well A1.
6. **Incubate for 30 min at room temperature (20...25 °C).**
7. Repeat step 4.
8. Dispense 50 µl Chikungunya virus Antibody Solution into all wells except for the Blank well A1.
9. **Incubate for 30 min at room temperature (20...25 °C).**
10. Repeat step 4.
11. Dispense 50 µl Streptavidin peroxidase conjugate into all wells except for the Blank well A1.
12. **Incubate for 30 min at room temperature (20...25 °C).** Do not expose to direct sunlight.
13. Repeat step 4.
14. Dispense 100 µl TMB solution into all wells
15. **Incubate for exact 15 min. at room temperature (20...25 °C) in the dark.** A blue colour occurs due to an enzymatic reaction.
16. Dispense 100 µl Stop Solution into all wells in the same order and at the same rate as for the TMB Substrate Solution, thereby a colour change from blue to yellow occurs.
17. Measure the absorbance at 450/620nm within 30 min after addition of the Stop Solution.

8.2. Measurement

Adjust the ELISA microwell plate reader **to zero** using the **Substrate Blank**.

If - due to technical reasons - the ELISA microwell plate reader cannot be adjusted to zero using the Substrate Blank, subtract its absorbance value from all other absorbance values measured in order to obtain reliable results!

Measure the absorbance of all wells at **450 nm** and record the absorbance values for each standard/control and sample in the-plate layout.

Bichromatic measurement using a reference wavelength of 620 nm is recommended.

Where applicable calculate the **mean absorbance values** of all duplicates.

9. RESULTS

9.1. Run Validation Criteria

In order for an assay to be considered valid, the following criteria must be met:

- **Substrate Blank:** Absorbance value < **0.100**
- **Negative Control:** Absorbance value < **Cut-off**
- **Cut-off Control:** Absorbance value **0.150 – 1.300**
- **Positive Control:** Absorbance value > **Cut-off**

If these criteria are not met, the test is not valid and must be repeated.

9.2. Calculation of Results

The Cut-off is the mean absorbance value of the Cut-off Control determinations.

Example: Absorbance value Cut-off control 0.44 + absorbance value Cut-off control 0.42 = 0.86 / 2 = 0.43

Cut-off = 0.43

9.2.1 Results in Units [U]

$$\frac{\text{Sample (mean) absorbance value} \times 10}{\text{Cut-off}} = [\text{Units} = \text{U}]$$

Example:
$$\frac{1.591 \times 10}{0.43} = 37 \text{ U}$$

9.3. Interpretation of Results

Cut-off	10 U	-
Positive	> 11 U	Antibodies against the pathogen are present. There has been a contact with the antigen (pathogen resp. vaccine).
Equivocal	9 – 11 U	Antibodies against the pathogen could not be detected clearly. It is recommended to repeat the test with a fresh sample in 2 to 4 weeks. If the result is equivocal again the sample is judged as negative .
Negative	< 9 U	The sample contains no antibodies against the pathogen. A previous contact with the antigen (pathogen resp. vaccine) is unlikely.

9.3.1. Antibody Isotypes and State of Infection

Serology	Significance
IgM	Characteristic of the primary antibody response High IgM titer with low IgG titer: → suggests a current or very recent infection Rare: → persisting IgM
IgG	Characteristic of the secondary antibody response May persist for several years High IgG titer with low IgM titer: → may indicate a past infection

10. SPECIFIC PERFORMANCE CHARACTERISTICS

The results refer to the groups of samples investigated; these are not guaranteed specifications.

10.1. Precision

Intraassay	n	Mean (OD)	CV (%)
#1	24	0.287	8.53
#2	24	0.769	5.82
#3	24	0.618	5.50
Interassay	n	Mean (U)	CV (%)
#1	12	32.42	4.88
#2	12	26.96	5.82
#3	12	5.95	12.84

10.2. Diagnostic Specificity

The diagnostic specificity is defined as the probability of the assay of scoring negative in the absence of the specific analyte. It is 100.0% (95% confidence interval: 95.01% - 100.0%).

10.3. Diagnostic Sensitivity

The diagnostic sensitivity is defined as the probability of the assay of scoring positive in the presence of the specific analyte. It is 100.0% (95% confidence interval: 96.19% - 100.0%).

10.4. Interferences

Interferences with hemolytic, lipemic or icteric samples are not observed up to a concentration of 10 mg/ml hemoglobin, 5 mg/ml triglycerides and 0.5 mg/ml bilirubin.

10.5. Cross Reactivity

Cross reactivity with antibodies against *Borrelia*, CMV and *Toxoplasma* cannot be excluded.

Interference with polyclonal stimulation of EBV infections is likely. In the presence of infectious Mononucleosis (Pfeiffer's Disease, EBV infection) polyclonal stimulation of B lymphocytes can occur. This may result in non-specific reactions in the detection of antibodies of the IgM class. Therefore it is recommended to exclude an EBV infection by differential diagnosis. Cross reactivity with antibodies against other alpha viruses cannot be excluded.

11. LIMITATIONS OF THE PROCEDURE

Bacterial contamination or repeated freeze-thaw cycles of the sample may affect the absorbance values.

12. PRECAUTIONS AND WARNINGS

- The test procedure, the information, the precautions and warnings in the instructions for use have to be strictly followed. The use of the test kits with analyzers and similar equipment has to be validated. Any change in design, composition and test procedure as well as for any use in combination with other products not approved by the manufacturer is not authorized; the user himself is responsible for such changes. The manufacturer is not liable for false results and incidents for these reasons. The manufacturer is not liable for any results by visual analysis of the samples.
- For research use only.
- All materials of human or animal origin should be regarded and handled as potentially infectious.
- All components of human origin used for the production of these reagents have been tested for anti-HIV antibodies, anti-HCV antibodies and HBsAg and have been found to be non-reactive.
- **The Chikungunya virus antigens are inactivated. All materials should still be regarded and handled as potentially infectious. Wear gloves while performing the test. We recommend using the antigen under BSL2 cabinet (clean bench).**
- Do not interchange reagents or strips of different production lots.
- No reagents of other manufacturers should be used along with reagents of this test kit.
- Do not use reagents after expiry date stated on the label.
- Use only clean pipette tips, dispensers, and lab ware.
- Do not interchange screw caps of reagent vials to avoid cross-contamination.
- Close reagent vials tightly immediately after use to avoid evaporation and microbial contamination.
- After first opening and subsequent storage check conjugate and standard/control vials for microbial contamination prior to further use.
- To avoid cross-contamination and falsely elevated results pipette samples and dispense reagents without splashing accurately into the wells.
- The ELISA is only designed for qualified personnel who are familiar with good laboratory practice.

12.1. Disposal Considerations

Residues of chemicals and preparations are generally considered as hazardous waste. The disposal of this kind of waste is regulated through national and regional laws and regulations. Contact your local authorities or waste management companies which will give advice on how to dispose hazardous waste.

13. ORDERING INFORMATION

Prod. No.: ELS61243 Chikungunya Virus IgM-μ-capture ELISA

BIBLIOGRAPHY

Bienz, Kurt A. (2005): Viruses as Human Pathogen. In Fritz H. Kayser, Kurt A. Bienz, Johannes Eckert, Rolf M. Zinkernagel: Medical microbiology. Stuttgart, New York: Thieme (Thieme Flexibook), pp. 412–474.

Hochedez, Patrick; Jaureguiberry, Stephane; Debruyne, Monique; Bossi, Philippe; Hausfater, Pierre; Brucker, Gilles et al. (2006): Chikungunya infection in travelers. In *Emerging infectious diseases* 12 (10), pp. 1565–1567. DOI: 10.3201/eid1210.060495.







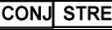






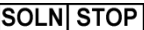



Markoff, Lewis (2005): Chapter 147: Alphaviruses. In Gerald L. Mandell, Robert Gordon Douglas, John Eugene Bennett (Eds.): Mandell, Douglas, and Bennett's principles and practice of infectious diseases, vol. 2. 6. ed. Philadelphia, Pa.: Elsevier, pp. 1913–1920.

Weaver, Scott C.; Tesh, Robert B.; Shope, Robert E. (2006): Alphavirus Infections. In Richard L. Guerrant, David H. Walker, Peter F. Weller (Eds.): Tropical infectious diseases. Principles, pathogens & practice. 2nd ed. Philadelphia: Churchill Livingstone, pp. 831–838.

ABBREVIATIONS

NMP	N-Methyl-2-pyrrolidone
-----	------------------------

SYMBOLS KEY

	Lot Number
	Expiration Date
	Storage Temperature
	Catalogue Number
	Consult Instructions for Use
	Microplate
	Streptavidin Conjugate
	Negative Control
	Positive Control
	Cut-off Control
	Antigen lyophilized
	Antibody solution
	Sample Diluent
	Stop solution
	TMB Substrate solution
	Washing Buffer 20x concentrated
	Contains sufficient for “n” tests

SCHEME OF THE ASSAY

Chikungunya Virus IgM μ -capture ELISA

Test Preparation

<p>Prepare reagents and samples as described.</p> <p>Establish the distribution and identification plan for all samples and standards/controls on the plate layout supplied in the kit.</p> <p>Select the required number of microtiter strips or wells and insert them into the holder.</p>
--

Assay Procedure

	Substrate Blank (A1)	Negative control	Cut-off control	Positive control	Sample (diluted 1+100)
Negative control	-	50 μ l	-	-	-
Cut-off control	-	-	50 μ l	-	-
Positive control	-	-	-	50 μ l	-
Sample (diluted 1+100)	-	-	-	-	50 μ l
Cover wells with foil supplied in the kit Incubate for 1 h at 37°C Wash each well three times with 300 μ l of Washing Buffer					
Reconstituted Antigen	-	50 μ l	50 μ l	50 μ l	50 μ l
Incubate for 30 min at room temperature (20...25 °C) Wash each well three times with 300 μ l of Washing Buffer					
Antibody Solution	-	50 μ l	50 μ l	50 μ l	50 μ l
Incubate for 30 min at room temperature (20...25 °C) Wash each well three times with 300 μ l of Washing Buffer					
Streptavidin conjugate	-	50 μ l	50 μ l	50 μ l	50 μ l
Incubate for 30 min at room temperature (20...25 °C) Do not expose to direct sunlight Wash each well three times with 300 μ l of Washing Buffer					
TMB Substrate solution	100 μ l	100 μ l	100 μ l	100 μ l	100 μ l
Incubate for exact 15 min at room temperature (20...25 °C) in the dark					
Stop solution	100 μ l	100 μ l	100 μ l	100 μ l	100 μ l
Photometric measurement at 450 nm (reference wavelength: 620 nm)					